

NOVEL HUMAN SEMAPHORIN AND
POLYNUCLEOTIDES ENCODING THE SAME

The present application claims the benefit of U.S.

5 Provisional Application Number 60/274,963 which was filed on
March 12, 2001 and is herein incorporated by reference in its
entirety.

1. INTRODUCTION

10 The present invention relates to the discovery,
identification, and characterization of novel human
polynucleotides encoding proteins that share sequence
similarity with mammalian semaphorin proteins. The invention
encompasses the described polynucleotides, host cell expression
15 systems, the encoded proteins, fusion proteins, polypeptides
and peptides, antibodies to the encoded proteins and peptides,
and genetically engineered animals that either lack or
overexpress the disclosed genes, antagonists and agonists of
the proteins, and other compounds that modulate the expression
20 or activity of the proteins encoded by the disclosed genes,
which can be used for diagnosis, drug screening, clinical trial
monitoring, the treatment of diseases and disorders, and
cosmetic or nutraceutical applications.

2. BACKGROUND OF THE INVENTION

25 Semaphorins are members of a superfamily of structurally
related proteins that have been associated with axon guidance.
Their expression in non-neural tissues indicate that
semaphorins can regulate biology in pathways beyond axon
guidance.

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3. SUMMARY OF THE INVENTION

The present invention relates to the discovery,
identification, and characterization of nucleotides that encode

novel human proteins, and the corresponding amino acid sequences of these proteins. The novel human semaphorin (NHSS) described for the first time herein share structural similarity with mammalian semaphorins and particularly semaphorin 6A1.

5 The novel human nucleic acid sequences described herein, encode alternative proteins/open reading frames (ORFs) of 1073, 998, 476, 1060, 985, and 463 amino acids in length (see respectively SEQ ID NOS: 2, 4, 6, 8, 10, and 12).

10 The invention also encompasses agonists and antagonists of the described NHSSs, including small molecules, large molecules, mutant NHSSs, or portions thereof, that compete with native NHS, peptides, and antibodies, as well as nucleotide sequences that can be used to inhibit the expression of the described NHSSs (e.g., antisense and ribozyme molecules, and open reading frame
15 or regulatory sequence replacement constructs) or to enhance the expression of the described NHSSs (e.g., expression constructs that place the described polynucleotide under the control of a strong promoter system), and transgenic animals that express a NHS sequence, or "knock-outs" (which can be
20 conditional) that do not express a functional NHS. Knock-out mice can be produced in several ways, one of which involves the use of mouse embryonic stem cells ("ES cells") lines that contain gene trap mutations in a murine homolog of at least one of the described NHSSs. When the unique NHS sequences described
25 in SEQ ID NOS:1-13 are "knocked-out" they provide a method of identifying phenotypic expression of the particular gene as well as a method of assigning function to previously unknown genes. In addition, animals in which the unique NHS sequences described in SEQ ID NOS:1-13 are "knocked-out" provide a unique
30 source in which to elicit antibodies to homologous and orthologous proteins which would have been previously viewed by the immune system as "self" and therefore would have failed to elicit significant antibody responses. To these ends, gene

trapped knockout ES cells have been generated in murine homologs of the described NHSs.

Additionally, the unique NHS sequences described in SEQ ID NOS:1-13 are useful for the identification of protein coding sequence and mapping a unique gene to a particular chromosome. These sequences identify actual, biologically verified, and therefore relevant, exon splice junctions as opposed to those that may have been bioinformatically predicted from genomic sequence alone. The sequences of the present invention are also useful as additional DNA markers for restriction fragment length polymorphism (RFLP) analysis, and in forensic biology.

Further, the present invention also relates to processes for identifying compounds that modulate, *i.e.*, act as agonists or antagonists, of NHS expression and/or NHS activity that utilize purified preparations of the described NHSs and/or NHS product, or cells expressing the same. Such compounds can be used as therapeutic agents for the treatment of any of a wide variety of symptoms associated with biological disorders or imbalances.

4. DESCRIPTION OF THE SEQUENCE LISTING AND FIGURES

The Sequence Listing provides the sequences of the described NHS ORFs that encode the described NHS amino acid sequences. SEQ ID NO:13 describes a NHS ORF and flanking sequences.

5. DETAILED DESCRIPTION OF THE INVENTION

The NHSs, described for the first time herein, are novel proteins that are expressed in, *inter alia*, human cell lines, human fetal brain, pituitary, cerebellum, spinal cord, lymph node, lung, kidney, prostate, thyroid, adrenal gland, small intestine, colon, skeletal muscle, uterus, placenta, mammary gland, skin, esophagus, bladder, pericardium, fetal kidney,

fetal lung, 6-, 9-, and 12-week embryos, and embryonic carcinoma cells.

The present invention encompasses the nucleotides presented in the Sequence Listing, host cells expressing such nucleotides, the expression products of such nucleotides, and:

5 (a) nucleotides that encode mammalian homologs of the described genes, including the specifically described NHSs, and the NHS products; (b) nucleotides that encode one or more portions of the NHSs that correspond to functional domains, and the

10 polypeptide products specified by such nucleotide sequences, including, but not limited to, the novel regions of any active domain(s); (c) isolated nucleotides that encode mutant versions, engineered or naturally occurring, of the described NHSs in which all or a part of at least one domain is deleted

15 or altered, and the polypeptide products specified by such nucleotide sequences, including, but not limited to, soluble proteins and peptides in which all or a portion of the signal sequence is deleted; (d) nucleotides that encode chimeric fusion proteins containing all or a portion of a coding region

20 of a NHS, or one of its domains (e.g., a receptor or ligand binding domain, accessory protein/self-association domain, etc.) fused to another peptide or polypeptide; or (e) therapeutic or diagnostic derivatives of the described polynucleotides such as oligonucleotides, antisense

25 polynucleotides, ribozymes, dsRNA, or gene therapy constructs comprising a sequence first disclosed in the Sequence Listing.

As discussed above, the present invention includes:

(a) the human DNA sequences presented in the Sequence Listing (and vectors comprising the same) and additionally contemplates

30 any nucleotide sequence encoding a contiguous NHS open reading frame (ORF) that hybridizes to a complement of a DNA sequence presented in the Sequence Listing under highly stringent conditions, e.g., hybridization to filter-bound DNA in 0.5 M

NaHPO₄, 7% sodium dodecyl sulfate (SDS), 1 mM EDTA at 65°C, and washing in 0.1xSSC/0.1% SDS at 68°C (Ausubel F.M. et al., eds., 1989, Current Protocols in Molecular Biology, Vol. I, Green Publishing Associates, Inc., and John Wiley & Sons, Inc., NY, at p. 2.10.3) and encodes a functionally equivalent expression product. Additionally contemplated are any nucleotide sequences that hybridize to the complement of a DNA sequence that encodes and expresses an amino acid sequence presented in the Sequence Listing under moderately stringent conditions, e.g., washing in 0.2xSSC/0.1% SDS at 42°C (Ausubel et al., 1989, *supra*), yet still encodes a functionally equivalent NHS product. Functional equivalents of a NHS include naturally occurring NHSs present in other species and mutant NHSs whether naturally occurring or engineered (by site directed mutagenesis, gene shuffling, directed evolution as described in, for example, U.S. Patent No. 5,837,458). The invention also includes degenerate nucleic acid variants of the disclosed NHS polynucleotide sequences.

Additionally contemplated are polynucleotides encoding NHS ORFs, or their functional equivalents, encoded by polynucleotide sequences that are about 99, 95, 90, or about 85 percent similar or identical to corresponding regions of the nucleotide sequences of the Sequence Listing (as measured by BLAST sequence comparison analysis using, for example, the GCG sequence analysis package (Madison, Wisconsin) using standard default settings).

The invention also includes nucleic acid molecules, preferably DNA molecules, that hybridize to, and are therefore the complements of, the described NHS gene nucleotide sequences. Such hybridization conditions may be highly stringent or less highly stringent, as described above. In instances where the nucleic acid molecules are deoxyoligonucleotides ("DNA oligos"), such molecules are

generally about 16 to about 100 bases long, or about 20 to about 80, or about 34 to about 45 bases long, or any variation or combination of sizes represented therein that incorporate a contiguous region of sequence first disclosed in the Sequence Listing. Such oligonucleotides can be used in conjunction with the polymerase chain reaction (PCR) to screen libraries, isolate clones, and prepare cloning and sequencing templates, etc.

Alternatively, such NHS oligonucleotides can be used as hybridization probes for screening libraries, and assessing gene expression patterns (particularly using a micro array or high-throughput "chip" format). Additionally, a series of the described NHS oligonucleotide sequences, or the complements thereof, can be used to represent all or a portion of the described NHS sequences. An oligonucleotide or polynucleotide sequence first disclosed in at least a portion of one or more of the sequences of SEQ ID NOS: 1-13 can be used as a hybridization probe in conjunction with a solid support matrix/substrate (resins, beads, membranes, plastics, polymers, metal or metallized substrates, crystalline or polycrystalline substrates, etc.). Of particular note are spatially addressable arrays (*i.e.*, gene chips, microtiter plates, etc.) of oligonucleotides and polynucleotides, or corresponding oligopeptides and polypeptides, wherein at least one of the biopolymers present on the spatially addressable array comprises an oligonucleotide or polynucleotide sequence first disclosed in at least one of the sequences of SEQ ID NOS: 1-13, or an amino acid sequence encoded thereby. Methods for attaching biopolymers to, or synthesizing biopolymers on, solid support matrices, and conducting binding studies thereon are disclosed in, *inter alia*, U.S. Patent Nos. 5,700,637, 5,556,752, 5,744,305, 4,631,211, 5,445,934, 5,252,743,

4,713,326, 5,424,186, and 4,689,405 the disclosures of which are herein incorporated by reference in their entirety.

Addressable arrays comprising sequences first disclosed in SEQ ID NOS:1-13 can be used to identify and characterize the temporal and tissue specific expression of a gene. These addressable arrays incorporate oligonucleotide sequences of sufficient length to confer the required specificity, yet be within the limitations of the production technology. The length of these probes is within a range of between about 8 to about 2000 nucleotides. Preferably the probes consist of 60 nucleotides and more preferably 25 nucleotides from the sequences first disclosed in SEQ ID NOS:1-13.

For example, a series of the described oligonucleotide sequences, or the complements thereof, can be used in chip format to represent all or a portion of the described sequences. The oligonucleotides, typically between about 16 to about 40 (or any whole number within the stated range) nucleotides in length can partially overlap each other and/or the sequence may be represented using oligonucleotides that do not overlap. Accordingly, the described polynucleotide sequences shall typically comprise at least about two or three distinct oligonucleotide sequences of at least about 8 nucleotides in length that are each first disclosed in the described Sequence Listing. Such oligonucleotide sequences can begin at any nucleotide present within a sequence in the Sequence Listing and proceed in either a sense (5'-to-3') orientation vis-a-vis the described sequence or in an antisense orientation.

Microarray-based analysis allows the discovery of broad patterns of genetic activity, providing new understanding of gene functions and generating novel and unexpected insight into transcriptional processes and biological mechanisms. The use of addressable arrays comprising sequences first disclosed in SEQ

ID NOS:1-13 provides detailed information about transcriptional changes involved in a specific pathway, potentially leading to the identification of novel components or gene functions that manifest themselves as novel phenotypes.

5 Probes consisting of sequences first disclosed in SEQ ID NOS:1-13 can also be used in the identification, selection and validation of novel molecular targets for drug discovery. The use of these unique sequences permits the direct confirmation of drug targets and recognition of drug dependent changes in
10 gene expression that are modulated through pathways distinct from the drugs intended target. These unique sequences therefore also have utility in defining and monitoring both drug action and toxicity.

As an example of utility, the sequences first disclosed in
15 SEQ ID NOS:1-13 can be utilized in microarrays or other assay formats, to screen collections of genetic material from patients who have a particular medical condition. These investigations can also be carried out using the sequences first disclosed in SEQ ID NOS:1-13 *in silico* and by comparing
20 previously collected genetic databases and the disclosed sequences using computer software known to those in the art.

Thus the sequences first disclosed in SEQ ID NOS:1-13 can be used to identify mutations associated with a particular disease and also as a diagnostic or prognostic assay.

25 Although the presently described sequences have been specifically described using nucleotide sequence, it should be appreciated that each of the sequences can uniquely be described using any of a wide variety of additional structural attributes, or combinations thereof. For example, a given
30 sequence can be described by the net composition of the nucleotides present within a given region of the sequence in conjunction with the presence of one or more specific oligonucleotide sequence(s) first disclosed in the SEQ ID NOS:

1-13. Alternatively, a restriction map specifying the relative positions of restriction endonuclease digestion sites, or various palindromic or other specific oligonucleotide sequences can be used to structurally describe a given sequence. Such
5 restriction maps, which are typically generated by widely available computer programs (e.g., the University of Wisconsin GCG sequence analysis package, SEQUENCHER 3.0, Gene Codes Corp., Ann Arbor, MI, etc.), can optionally be used in conjunction with one or more discrete nucleotide sequence(s)
10 present in the sequence that can be described by the relative position of the sequence relative to one or more additional sequence(s) or one or more restriction sites present in the disclosed sequence.

For oligonucleotide probes, highly stringent conditions
15 may refer, e.g., to washing in 6x SSC/0.05% sodium pyrophosphate at 37°C (for 14-base oligos), 48°C (for 17-base oligos), 55°C (for 20-base oligos), and 60°C (for 23-base oligos). These nucleic acid molecules may encode or act as NHS gene antisense molecules, useful, for example, in NHS gene
20 regulation and/or as antisense primers in amplification reactions of NHS gene nucleic acid sequences. With respect to NHS gene regulation, such techniques can be used to regulate biological functions. Further, such sequences may be used as part of ribozyme and/or triple helix sequences that are also
25 useful for NHS gene regulation.

Inhibitory antisense or double stranded oligonucleotides can additionally comprise at least one modified base moiety which is selected from the group including, but not limited to, 5-fluorouracil, 5-bromouracil, 5-chlorouracil, 5-iodouracil,
30 hypoxanthine, xanthine, 4-acetylcytosine, 5-(carboxyhydroxymethyl) uracil, 5-carboxymethylaminomethyl-2-thiouridine, 5-carboxymethylaminomethyluracil, dihydrouracil, beta-D-galactosylqueosine, inosine, N6-isopentenyladenine,

1-methylguanine, 1-methylinosine, 2,2-dimethylguanine,
2-methyladenine, 2-methylguanine, 3-methylcytosine,
5-methylcytosine, N6-adenine, 7-methylguanine,
5-methylaminomethyluracil, 5-methoxyaminomethyl-2-thiouracil,
5 beta-D-mannosylqueosine, 5'-methoxycarboxymethyluracil,
5-methoxyuracil, 2-methylthio-N6-isopentenyladenine, uracil-5-
oxyacetic acid (v), wybutoxosine, pseudouracil, queosine,
2-thiocytosine, 5-methyl-2-thiouracil, 2-thiouracil,
4-thiouracil, 5-methyluracil, uracil-5-oxyacetic acid
10 methylester, uracil-5-oxyacetic acid (v), 5-methyl-
2-thiouracil, 3-(3-amino-3-N-2-carboxypropyl) uracil, (acp3)w,
and 2,6-diaminopurine.

The antisense oligonucleotide can also comprise at least
one modified sugar moiety selected from the group including,
15 but not limited to, arabinose, 2-fluoroarabinose, xylulose, and
hexose.

In yet another embodiment, the antisense oligonucleotide
will comprise at least one modified phosphate backbone selected
from the group including, but not limited to, a
20 phosphorothioate, a phosphorodithioate, a phosphoramidothioate,
a phosphoramidate, a phosphordiamidate, a methylphosphonate, an
alkyl phosphotriester, and a formacetal or analog thereof.

In yet another embodiment, the antisense oligonucleotide
is an α -anomeric oligonucleotide. An α -anomeric
25 oligonucleotide forms specific double-stranded hybrids with
complementary RNA in which, contrary to the usual β -units, the
strands run parallel to each other (Gautier *et al.*, 1987, Nucl.
Acids Res. 15:6625-6641). The oligonucleotide is a 2'-O-
methylribonucleotide (Inoue *et al.*, 1987, Nucl. Acids Res.
30 15:6131-6148), or a chimeric RNA-DNA analogue (Inoue *et al.*,
1987, FEBS Lett. 215:327-330). Alternatively, double stranded
RNA can be used to disrupt the expression and function of a
targeted NHS.

Oligonucleotides of the invention can be synthesized by standard methods known in the art, e.g. by use of an automated DNA synthesizer (such as are commercially available from Biosearch, Applied Biosystems, etc.). As examples,

5 phosphorothioate oligonucleotides can be synthesized by the method of Stein et al. (1988, Nucl. Acids Res. 16:3209), and methylphosphonate oligonucleotides can be prepared by use of controlled pore glass polymer supports (Sarin et al., 1988, Proc. Natl. Acad. Sci. USA. 85:7448-7451), etc.

10 Low stringency conditions are well-known to those of skill in the art, and will vary predictably depending on the specific organisms from which the library and the labeled sequences are derived. For guidance regarding such conditions see, for example, Sambrook et al., 1989, Molecular Cloning, A Laboratory
15 Manual (and periodic updates thereof), Cold Spring Harbor Press, NY; and Ausubel et al., 1989, Current Protocols in Molecular Biology, Green Publishing Associates and Wiley Interscience, NY.

Alternatively, suitably labeled NHS nucleotide probes can
20 be used to screen a human genomic library using appropriately stringent conditions or by PCR. The identification and characterization of human genomic clones is helpful for identifying polymorphisms (including, but not limited to, nucleotide repeats, microsatellite alleles, single nucleotide
25 polymorphisms, or coding single nucleotide polymorphisms), determining the genomic structure of a given locus/allele, and designing diagnostic tests. For example, sequences derived from regions adjacent to the intron/exon boundaries of the human gene can be used to design primers for use in
30 amplification assays to detect mutations within the exons, introns, splice sites (e.g., splice acceptor and/or donor sites), etc., that can be used in diagnostics and pharmacogenomics.

In another example, the present sequences can be used in restriction fragment length polymorphism (RFLP) analysis to identify specific individuals. In this technique, an individual's genomic DNA is digested with one or more
5 restriction enzymes, and probed on a Southern blot to yield unique bands for identification (as generally described in U.S. Patent No. 5,272,057, incorporated herein by reference). In addition, the sequences of the present invention can be used to provide polynucleotide reagents, e.g., PCR primers, targeted to
10 specific loci in the human genome, which can enhance the reliability of DNA-based forensic identifications by, for example, providing another "identification marker" (i.e., another DNA sequence that is unique to a particular individual). Actual base sequence information can be used for
15 identification as an accurate alternative to patterns formed by restriction enzyme generated fragments.

Further, a NHS gene homolog can be isolated from nucleic acid from an organism of interest by performing PCR using two degenerate or "wobble" oligonucleotide primer pools designed on
20 the basis of amino acid sequences within the NHS products disclosed herein. The template for the reaction may be total RNA, mRNA, and/or cDNA obtained by reverse transcription of mRNA prepared from human or non-human cell lines or tissue known or suspected to express an allele of a NHS gene.

25 The PCR product can be subcloned and sequenced to ensure that the amplified sequences represent the sequence of the desired NHS gene. The PCR fragment can then be used to isolate a full length cDNA clone by a variety of methods. For example, the amplified fragment can be labeled and used to screen a cDNA
30 library, such as a bacteriophage cDNA library. Alternatively, the labeled fragment can be used to isolate genomic clones via the screening of a genomic library.

PCR technology can also be used to isolate full length cDNA sequences. For example, RNA can be isolated, following standard procedures, from an appropriate cellular or tissue source (i.e., one known, or suspected, to express a NHS gene).

5 A reverse transcription (RT) reaction can be performed on the RNA using an oligonucleotide primer specific for the most 5' end of the amplified fragment for the priming of first strand synthesis. The resulting RNA/DNA hybrid may then be "tailed" using a standard terminal transferase reaction, the hybrid may
10 be digested with RNase H, and second strand synthesis may then be primed with a complementary primer. Thus, cDNA sequences upstream of the amplified fragment can be isolated. For a review of cloning strategies that can be used, see e.g., Sambrook et al., 1989, *supra*.

15 A cDNA encoding a mutant NHS sequence can be isolated, for example, by using PCR. In this case, the first cDNA strand may be synthesized by hybridizing an oligo-dT oligonucleotide to mRNA isolated from tissue known or suspected to be expressed in an individual putatively carrying a mutant NHS allele, and by
20 extending the new strand with reverse transcriptase. The second strand of the cDNA is then synthesized using an oligonucleotide that hybridizes specifically to the 5' end of the normal sequence. Using these two primers, the product is then amplified via PCR, optionally cloned into a suitable
25 vector, and subjected to DNA sequence analysis through methods well-known to those of skill in the art. By comparing the DNA sequence of the mutant NHS allele to that of a corresponding normal NHS allele, the mutation(s) responsible for the loss or alteration of function of the mutant NHS gene product can be
30 ascertained.

Alternatively, a genomic library can be constructed using DNA obtained from an individual suspected of or known to carry a mutant NHS allele (e.g., a person manifesting a NHS-

associated phenotype such as, for example, obesity, behavioral disorders, colitis or spastic colon, high blood pressure, depression, infertility, etc.), or a cDNA library can be constructed using RNA from a tissue known, or suspected, to express a mutant NHS allele. A normal NHS gene, or any suitable fragment thereof, can then be labeled and used as a probe to identify the corresponding mutant NHS allele in such libraries. Clones containing mutant NHS sequences can then be purified and subjected to sequence analysis according to methods well-known to those skilled in the art.

Additionally, an expression library can be constructed utilizing cDNA synthesized from, for example, RNA isolated from a tissue known, or suspected, to express a mutant NHS allele in an individual suspected of or known to carry such a mutant allele. In this manner, gene products made by the putatively mutant tissue can be expressed and screened using standard antibody screening techniques in conjunction with antibodies raised against a normal NHS product, as described below. For screening techniques, see, for example, Harlow, E. and Lane, eds., 1988, "Antibodies: A Laboratory Manual", Cold Spring Harbor Press, Cold Spring Harbor, NY.

Additionally, screening can be accomplished by screening with labeled NHS fusion proteins, such as, for example, alkaline phosphatase-NHS or NHS-alkaline phosphatase fusion proteins. In cases where a NHS mutation results in an expression product with altered function (e.g., as a result of a missense or a frameshift mutation), polyclonal antibodies to NHS are likely to cross-react with a corresponding mutant NHS expression product. Library clones detected via their reaction with such labeled antibodies can be purified and subjected to sequence analysis according to methods well-known in the art.

The invention also encompasses (a) DNA vectors that contain any of the foregoing NHS coding sequences and/or their

complements (*i.e.*, antisense); (b) DNA expression vectors that contain any of the foregoing NHS coding sequences operatively associated with a regulatory element that directs the expression of the coding sequences (for example, baculovirus as described in U.S. Patent No. 5,869,336 herein incorporated by reference); (c) genetically engineered host cells that contain any of the foregoing NHS coding sequences operatively associated with a regulatory element that directs the expression of the coding sequences in the host cell; and (d) genetically engineered host cells that express an endogenous NHS sequence under the control of an exogenously introduced regulatory element (*i.e.*, gene activation). As used herein, regulatory elements include, but are not limited to, inducible and non-inducible promoters, enhancers, operators and other elements known to those skilled in the art that drive and regulate expression. Such regulatory elements include, but are not limited to, the cytomegalovirus (hCMV) immediate early gene, regulatable, viral elements (particularly retroviral LTR promoters), the early or late promoters of SV40 adenovirus, the *lac* system, the *trp* system, the *TAC* system, the *TRC* system, the major operator and promoter regions of phage lambda, the control regions of fd coat protein, the promoter for 3-phosphoglycerate kinase (PGK), the promoters of acid phosphatase, and the promoters of the yeast α -mating factors.

The present invention also encompasses antibodies and anti-idiotypic antibodies (including Fab fragments), antagonists and agonists of a NHS, as well as compounds or nucleotide constructs that inhibit expression of a NHS sequence (transcription factor inhibitors, antisense and ribozyme molecules, or open reading frame sequence or regulatory sequence replacement constructs), or promote the expression of a NHS (*e.g.*, expression constructs in which NHS coding

sequences are operatively associated with expression control elements such as promoters, promoter/enhancers, etc.).

The NHSSs or NHS peptides, NHS fusion proteins, NHS nucleotide sequences, antibodies, antagonists and agonists can be useful for the detection of mutant NHSSs or inappropriately expressed NHSSs for the diagnosis of disease. The NHS proteins or peptides, NHS fusion proteins, NHS nucleotide sequences, host cell expression systems, antibodies, antagonists, agonists and genetically engineered cells and animals can be used for screening for drugs (or high throughput screening of combinatorial libraries) effective in the treatment of the symptomatic or phenotypic manifestations of perturbing the normal function of NHS in the body. The use of engineered host cells and/or animals may offer an advantage in that such systems allow not only for the identification of compounds that bind to the endogenous receptor for an NHS, but can also identify compounds that trigger NHS-mediated activities or pathways.

Finally, the NHS products, or modified/processed forms thereof, can be used as therapeutics. For example, soluble derivatives such as NHS peptides/domains corresponding to the NHSSs, NHS fusion protein products (especially NHS-Ig fusion proteins, i.e., fusions of a NHS, or a domain of a NHS, to an IgFc), NHS antibodies and anti-idiotypic antibodies (including Fab fragments), antagonists or agonists (including compounds that modulate or act on downstream targets in a NHS-mediated pathway) can be used to directly treat diseases or disorders. For instance, the administration of an effective amount of soluble NHS, or a NHS-IgFc fusion protein or an anti-idiotypic antibody (or its Fab) that mimics the NHS could activate or effectively antagonize the endogenous NHS receptor. Nucleotide constructs encoding such NHS products can be used to genetically engineer host cells to express such products in

vivo; these genetically engineered cells function as "bioreactors" in the body delivering a continuous supply of a NHS, a NHS peptide, or a NHS fusion protein to the body. Nucleotide constructs encoding functional NHSs, mutant NHSs, as well as antisense and ribozyme molecules can also be used in "gene therapy" approaches for the modulation of NHS expression. Thus, the invention also encompasses pharmaceutical formulations and methods for treating biological disorders such as, for example, hyperthyroidism and/or hypothyroidism.

Various aspects of the invention are described in greater detail in the subsections below.

5.1 THE NHS SEQUENCES

The cDNA sequences and the corresponding deduced amino acid sequences of the described NHSs are presented in the Sequence Listing. The NHS nucleotides were obtained from clustered human gene trapped sequences, and cDNA products isolated from human skeletal muscle, colon, small intestine, fetal kidney, fetus, fetal brain, fetal lung, and mammary gland mRNAs (Clontech, Palo Alto, CA). The described sequences share substantial structural similarity with a variety of proteins, including, but not limited to, semaphorins. Because of their potential medical significance, semaphorin protein family members have been subject to considerable scientific scrutiny as evidenced in U.S. Patent Nos. 5,981,222 and 6,013,781, which are herein incorporated by reference, and which describe various applications, uses, and compositions in which the presently described NHSs can be advantageously applied.

The described NHS sequences can contain a variety of polymorphisms such as a T/A polymorphism that can occur in the sequence region represented by nucleotide position number 1108 of, for example, SEQ ID NO:1 that can give rise to a cys or ser being present at corresponding amino acid positions 370 of, for

example, SEQ ID NO:2, a G/A polymorphism at nucleotide position 1433 of, for example, SEQ ID NO:1 which can give rise to a ser or asn being present at corresponding amino acid position 478 of, for example, SEQ ID NO:2, and a A/G polymorphism at
5 nucleotides position number 2366 of, for example, SEQ ID NO:1 which can result in a lys or arg being present at corresponding amino acid position number 789 of, for example, SEQ ID NO:2. The present invention contemplates sequences comprising any of the above polymorphisms and all combinations and permutations
10 thereof.

The described sequences are apparently encoded on human chromosome 15 (see GENBANK accession no. AC018900 and AK021660). Accordingly, the described sequences can be used to map the corresponding coding regions of the human genome.

15 The described novel human polynucleotide sequences can be used, among other things, in the molecular mutagenesis/evolution of proteins that are at least partially encoded by the described novel sequences using, for example, polynucleotide shuffling or related methodologies. Such
20 approaches are described in U.S. Patent Nos. 5,830,721 and 5,837,458 which are herein incorporated by reference in their entirety.

NHS gene products can also be expressed in transgenic animals. Animals of any species, including, but not limited
25 to, worms, mice, rats, rabbits, guinea pigs, pigs, micro-pigs, birds, goats, and non-human primates, e.g., baboons, monkeys, and chimpanzees may be used to generate NHS transgenic animals.

Any technique known in the art may be used to introduce a NHS transgene into animals to produce the founder lines of
30 transgenic animals. Such techniques include, but are not limited to, pronuclear microinjection (Hoppe, P.C. and Wagner, T.E., 1989, U.S. Patent No. 4,873,191); retrovirus-mediated gene transfer into germ lines (Van der Putten et al., 1985,

Proc. Natl. Acad. Sci. USA 82:6148-6152); gene targeting in embryonic stem cells (Thompson *et al.*, 1989, Cell 56:313-321); electroporation of embryos (Lo, 1983, Mol Cell. Biol. 3:1803-1814); and sperm-mediated gene transfer (Lavitrano *et al.*, 1989, Cell 57:717-723); *etc.* For a review of such techniques, see Gordon, 1989, Transgenic Animals, Intl. Rev. Cytol. 115:171-229, which is incorporated by reference herein in its entirety.

The present invention provides for transgenic animals that carry the NHS transgene in all their cells, as well as animals which carry the transgene in some, but not all their cells, *i.e.*, mosaic animals or somatic cell transgenic animals. The transgene may be integrated as a single transgene or in concatamers, *e.g.*, head-to-head tandems or head-to-tail tandems. The transgene may also be selectively introduced into and activated in a particular cell-type by following, for example, the teaching of Lasko *et al.*, 1992, Proc. Natl. Acad. Sci. USA 89:6232-6236. The regulatory sequences required for such a cell-type specific activation will depend upon the particular cell-type of interest, and will be apparent to those of skill in the art.

When it is desired that a NHS transgene be integrated into the chromosomal site of the endogenous NHS gene, gene targeting is preferred. Briefly, when such a technique is to be utilized, vectors containing some nucleotide sequences homologous to the endogenous NHS gene are designed for the purpose of integrating, via homologous recombination with chromosomal sequences, into and disrupting the function of the nucleotide sequence of the endogenous NHS gene (*i.e.*, "knockout" animals).

The transgene can also be selectively introduced into a particular cell-type, thus inactivating the endogenous NHS gene in only that cell-type, by following, for example, the teaching

of Gu et al., 1994, Science, 265:103-106. The regulatory sequences required for such a cell-type specific inactivation will depend upon the particular cell-type of interest, and will be apparent to those of skill in the art.

5 Once transgenic animals have been generated, the expression of the recombinant NHS gene may be assayed utilizing standard techniques. Initial screening may be accomplished by Southern blot analysis or PCR techniques to analyze animal tissues to assay whether integration of the transgene has taken
10 place. The level of mRNA expression of the transgene in the tissues of the transgenic animals may also be assessed using techniques which include, but are not limited to, Northern blot analysis of tissue samples obtained from the animal, *in situ* hybridization analysis, and RT-PCR. Samples of NHS gene-
15 expressing tissue, may also be evaluated immunocytochemically using antibodies specific for the NHS transgene product.

 The present invention provides for "knockin" animals. Knockin animals are those in which a gene that the animal does not naturally have in its genome, is inserted. For example,
20 when a human gene is used to replace its murine ortholog in the mouse. Such knockin animals are useful for the *in vivo* study, testing and validation of, *intra alia*, human drug targets as well as for compounds that are directed at the same.

25 5.2 NHSS AND NHS POLYPEPTIDES

 NHSSs, polypeptides, peptide fragments, mutated, truncated, or deleted forms of the NHSSs, and/or NHS fusion proteins can be prepared for a variety of uses. These uses include, but are not limited to, use as protein therapeutics, the generation of
30 antibodies, as reagents in diagnostic assays, the identification of other cellular gene products related to a NHS, as reagents in assays for screening for compounds that can be used as pharmaceutical reagents useful in the therapeutic

treatment of mental, biological, or medical disorders and diseases. Given the similarity information and expression data, the described NHSs can be targeted (by drugs, oligos, antibodies, etc,) in order to treat disease, or to

5 therapeutically augment the efficacy of therapeutic agents.

The Sequence Listing discloses the amino acid sequences encoded by the described NHS sequences. The NHSs display initiator methionines in DNA sequence contexts consistent with a translation initiation site, and a signal sequence
10 characteristic of secreted or membrane proteins.

The NHS amino acid sequences of the invention include the amino acid sequences presented in the Sequence Listing as well as analogues and derivatives thereof. Further, corresponding NHS homologues from other species are encompassed by the
15 invention. In fact, any NHS protein encoded by the NHS nucleotide sequences described above are within the scope of the invention, as are any novel polynucleotide sequences encoding all or any novel portion of an amino acid sequence presented in the Sequence Listing. The degenerate nature of
20 the genetic code is well-known, and, accordingly, each amino acid presented in the Sequence Listing, is generically representative of the well-known nucleic acid "triplet" codon, or in many cases codons, that can encode the amino acid. As such, as contemplated herein, the amino acid sequences
25 presented in the Sequence Listing, when taken together with the genetic code (see, for example, Table 4-1 at page 109 of "Molecular Cell Biology", 1986, J. Darnell et al. eds., Scientific American Books, New York, NY, herein incorporated by reference) are generically representative of all the various
30 permutations and combinations of nucleic acid sequences that can encode such amino acid sequences.

The invention also encompasses proteins that are functionally equivalent to the NHSs encoded by the presently

described nucleotide sequences as judged by any of a number of criteria, including, but not limited to, the ability to bind and cleave a substrate of a NHS, or the ability to effect an identical or complementary downstream pathway, or a change in cellular metabolism (e.g., proteolytic activity, ion flux, tyrosine phosphorylation, transport, etc.). Such functionally equivalent NHS proteins include, but are not limited to, additions or substitutions of amino acid residues within the amino acid sequence encoded by the NHS nucleotide sequences described above, but which result in a silent change, thus producing a functionally equivalent expression product. Amino acid substitutions may be made on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues involved. For example, nonpolar (hydrophobic) amino acids include alanine, leucine, isoleucine, valine, proline, phenylalanine, tryptophan, and methionine; polar neutral amino acids include glycine, serine, threonine, cysteine, tyrosine, asparagine, and glutamine; positively charged (basic) amino acids include arginine, lysine, and histidine; and negatively charged (acidic) amino acids include aspartic acid and glutamic acid.

A variety of host-expression vector systems can be used to express the NHS nucleotide sequences of the invention. Where, as in the present instance, the NHS peptide or polypeptide is thought to be a secreted protein, the hydrophobic regions of the protein can be excised and the resulting soluble peptide or polypeptide can be recovered from the culture media. Such expression systems also encompass engineered host cells that express a NHS, or functional equivalent, *in situ*. Purification or enrichment of a NHS from such expression systems can be accomplished using appropriate detergents and lipid micelles and methods well-known to those skilled in the art. However, such engineered host cells themselves may be used in situations

where it is important not only to retain the structural and functional characteristics of the NHS, but to assess biological activity, e.g., in certain drug screening assays.

The expression systems that may be used for purposes of the invention include, but are not limited to, microorganisms such as bacteria (e.g., *E. coli*, *B. subtilis*) transformed with recombinant bacteriophage DNA, plasmid DNA or cosmid DNA expression vectors containing NHS nucleotide sequences; yeast (e.g., *Saccharomyces*, *Pichia*) transformed with recombinant yeast expression vectors containing NHS nucleotide sequences; insect cell systems infected with recombinant virus expression vectors (e.g., baculovirus) containing NHS nucleotide sequences; plant cell systems infected with recombinant virus expression vectors (e.g., cauliflower mosaic virus, CaMV; tobacco mosaic virus, TMV) or transformed with recombinant plasmid expression vectors (e.g., Ti plasmid) containing NHS nucleotide sequences; or mammalian cell systems (e.g., COS, CHO, BHK, 293, 3T3) harboring recombinant expression constructs containing NHS nucleotide sequences and promoters derived from the genome of mammalian cells (e.g., metallothionein promoter) or from mammalian viruses (e.g., the adenovirus late promoter; the vaccinia virus 7.5K promoter).

In bacterial systems, a number of expression vectors may be advantageously selected depending upon the use intended for the NHS product being expressed. For example, when a large quantity of such a protein is to be produced for the generation of pharmaceutical compositions of or containing NHS, or for raising antibodies to a NHS, vectors that direct the expression of high levels of fusion protein products that are readily purified may be desirable. Such vectors include, but are not limited, to the *E. coli* expression vector pUR278 (Ruther et al., 1983, EMBO J. 2:1791), in which a NHS coding sequence may be ligated individually into the vector in frame with the *lacZ*

coding region so that a fusion protein is produced; pIN vectors
(Inouye & Inouye, 1985, Nucleic Acids Res. 13:3101-3109; Van
Heeke & Schuster, 1989, J. Biol. Chem. 264:5503-5509); and the
like. pGEX vectors (Pharmacia or American Type Culture
5 Collection) can also be used to express foreign polypeptides as
fusion proteins with glutathione S-transferase (GST). In
general, such fusion proteins are soluble and can easily be
purified from lysed cells by adsorption to glutathione-agarose
beads followed by elution in the presence of free glutathione.
10 The PGEX vectors are designed to include thrombin or factor Xa
protease cleavage sites so that the cloned target expression
product can be released from the GST moiety.

In an insect system, *Autographa californica* nuclear
polyhedrosis virus (AcNPV) is used as a vector to express
15 foreign polynucleotide sequences. The virus grows in
Spodoptera frugiperda cells. A NHS coding sequence can be
cloned individually into non-essential regions (for example the
polyhedrin gene) of the virus and placed under control of an
AcNPV promoter (for example the polyhedrin promoter).
20 Successful insertion of NHS coding sequence will result in
inactivation of the polyhedrin gene and production of non-
occluded recombinant virus (i.e., virus lacking the
proteinaceous coat coded for by the polyhedrin gene). These
recombinant viruses are then used to infect *Spodoptera*
25 *frugiperda* cells in which the inserted sequence is expressed
(e.g., see Smith et al., 1983, J. Virol. 46: 584; Smith, U.S.
Patent No. 4,215,051).

In mammalian host cells, a number of viral-based
expression systems may be utilized. In cases where an
30 adenovirus is used as an expression vector, the NHS nucleotide
sequence of interest may be ligated to an adenovirus
transcription/translation control complex, e.g., the late
promoter and tripartite leader sequence. This chimeric

sequence may then be inserted in the adenovirus genome by *in vitro* or *in vivo* recombination. Insertion in a non-essential region of the viral genome (e.g., region E1 or E3) will result in a recombinant virus that is viable and capable of expressing a NHS product in infected hosts (e.g., See Logan & Shenk, 1984, Proc. Natl. Acad. Sci. USA 81:3655-3659). Specific initiation signals may also be required for efficient translation of inserted NHS nucleotide sequences. These signals include the ATG initiation codon and adjacent sequences. In cases where an entire NHS gene or cDNA, including its own initiation codon and adjacent sequences, is inserted into the appropriate expression vector, no additional translational control signals may be needed. However, in cases where only a portion of a NHS coding sequence is inserted, exogenous translational control signals, including, perhaps, the ATG initiation codon, must be provided. Furthermore, the initiation codon must be in phase with the reading frame of the desired coding sequence to ensure translation of the entire insert. These exogenous translational control signals and initiation codons can be of a variety of origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of appropriate transcription enhancer elements, transcription terminators, etc. (See Bitter *et al.*, 1987, Methods in Enzymol. 153:516-544).

In addition, a host cell strain may be chosen that modulates the expression of the inserted sequences, or modifies and processes the expression product in the specific fashion desired. Such modifications (e.g., glycosylation) and processing (e.g., cleavage) of protein products may be important for the function of the protein. Different host cells have characteristic and specific mechanisms for the post-translational processing and modification of proteins and expression products. Appropriate cell lines or host systems

can be chosen to ensure the correct modification and processing of the foreign protein expressed. To this end, eukaryotic host cells which possess the cellular machinery for proper processing of the primary transcript, glycosylation, and

5 phosphorylation of the expression product may be used. Such mammalian host cells include, but are not limited to, CHO, VERO, BHK, HeLa, COS, MDCK, 293, 3T3, WI38, and in particular, human cell lines.

For long-term, high-yield production of recombinant
10 proteins, stable expression is preferred. For example, cell lines which stably express the NHS sequences described above can be engineered. Rather than using expression vectors which contain viral origins of replication, host cells can be transformed with DNA controlled by appropriate expression
15 control elements (e.g., promoter, enhancer sequences, transcription terminators, polyadenylation sites, etc.), and a selectable marker. Following the introduction of the foreign DNA, engineered cells may be allowed to grow for 1-2 days in an enriched media, and then are switched to a selective media.

20 The selectable marker in the recombinant plasmid confers resistance to the selection and allows cells to stably integrate the plasmid into their chromosomes and grow to form foci which in turn can be cloned and expanded into cell lines. This method may advantageously be used to engineer cell lines
25 which express the NHS product. Such engineered cell lines may be particularly useful in screening and evaluation of compounds that affect the endogenous activity of the NHS product.

A number of selection systems may be used, including, but not limited to, the herpes simplex virus thymidine kinase
30 (Wigler et al., 1977, Cell 11:223), hypoxanthine-guanine phosphoribosyltransferase (Szybalska and Szybalski, 1962, Proc. Natl. Acad. Sci. USA 48:2026), and adenine phosphoribosyltransferase (Lowy et al., 1980, Cell 22:817)

genes, which can be employed in tk⁻, hgp⁻ or ap⁻ cells, respectively. Also, antimetabolite resistance can be used as the basis of selection for the following genes: dhfr, which confers resistance to methotrexate (Wigler *et al.*, 1980, Proc. Natl. Acad. Sci. USA 77:3567; O'Hare *et al.*, 1981, Proc. Natl. Acad. Sci. USA 78:1527); gpt, which confers resistance to mycophenolic acid (Mulligan and Berg, 1981, Proc. Natl. Acad. Sci. USA 78:2072); neo, which confers resistance to the aminoglycoside G-418 (Colberre-Garapin *et al.*, 1981, J. Mol. Biol. 150:1); and hyg⁺, which confers resistance to hygromycin (Santerre *et al.*, 1984, Gene 30:147).

Alternatively, any fusion protein can be readily purified by utilizing an antibody specific for the fusion protein being expressed. For example, a system described by Janknecht *et al.* allows for the ready purification of non-denatured fusion proteins expressed in human cell lines (Janknecht, *et al.*, 1991, Proc. Natl. Acad. Sci. USA 88:8972-8976). In this system, the sequence of interest is subcloned into a vaccinia recombination plasmid such that the sequence's open reading frame is translationally fused to an amino-terminal tag consisting of six histidine residues. Extracts from cells infected with recombinant vaccinia virus are loaded onto Ni²⁺-nitriloacetic acid-agarose columns and histidine-tagged proteins are selectively eluted with imidazole-containing buffers.

Also encompassed by the present invention are fusion proteins that direct the NHS to a target organ and/or facilitate transport across the membrane into the cytosol. Conjugation of NHSs to antibody molecules or their Fab fragments could be used to target cells bearing a particular epitope. Attaching the appropriate signal sequence to the NHS would also transport the NHS to the desired location within the cell. Alternatively targeting of NHS or its nucleic acid

sequence might be achieved using liposome or lipid complex based delivery systems. Such technologies are described in "Liposomes: A Practical Approach", New, R.R.C., ed., Oxford University Press, New York and in U.S. Patent Nos. 4,594,595, 5,459,127, 5,948,767 and 6,110,490 and their respective disclosures which are herein incorporated by reference in their entirety. Additionally embodied are novel protein constructs engineered in such a way that they facilitate transport of the NHS to the target site or desired organ, where they cross the cell membrane and/or the nucleus where the NHS can exert its functional activity. This goal may be achieved by coupling of the NHS to a cytokine or other ligand that provides targeting specificity, and/or to a protein transducing domain (see generally U.S. applications Ser. No. 60/111,701 and 60/056,713, both of which are herein incorporated by reference, for examples of such transducing sequences) to facilitate passage across cellular membranes and can optionally be engineered to include nuclear localization.

5.3 ANTIBODIES TO NHS PRODUCTS

Antibodies that specifically recognize one or more epitopes of a NHS, or epitopes of conserved variants of a NHS, or peptide fragments of a NHS are also encompassed by the invention. Such antibodies include, but are not limited to, polyclonal antibodies, monoclonal antibodies (mAbs), humanized or chimeric antibodies, single chain antibodies, Fab fragments, F(ab')₂ fragments, fragments produced by a Fab expression library, anti-idiotypic (anti-Id) antibodies, and epitope-binding fragments of any of the above.

The antibodies of the invention may be used, for example, in the detection of NHS in a biological sample and may, therefore, be utilized as part of a diagnostic or prognostic technique whereby patients may be tested for abnormal amounts

of NHS. Such antibodies may also be utilized in conjunction with, for example, compound screening schemes for the evaluation of the effect of test compounds on expression and/or activity of a NHS expression product. Additionally, such

5 antibodies can be used in conjunction gene therapy to, for example, evaluate the normal and/or engineered NHS-expressing cells prior to their introduction into the patient. Such antibodies may additionally be used as a method for the inhibition of abnormal NHS activity. Thus, such antibodies
10 may, therefore, be utilized as part of treatment methods.

For the production of antibodies, various host animals may be immunized by injection with the NHS, an NHS peptide (e.g., one corresponding to a functional domain of an NHS), truncated NHS polypeptides (NHS in which one or more domains have been
15 deleted), functional equivalents of the NHS or mutated variant of the NHS. Such host animals may include, but are not limited to, pigs, rabbits, mice, goats, and rats, to name but a few. Various adjuvants may be used to increase the immunological response, depending on the host species, including, but not
20 limited to, Freund's adjuvant (complete and incomplete), mineral salts such as aluminum hydroxide or aluminum phosphate, chitosan, surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, and potentially useful human adjuvants such as BCG (bacille
25 Calmette-Guerin) and *Corynebacterium parvum*. Alternatively, the immune response could be enhanced by combination and or coupling with molecules such as keyhole limpet hemocyanin, tetanus toxoid, diphtheria toxoid, ovalbumin, cholera toxin or fragments thereof. Polyclonal antibodies are heterogeneous
30 populations of antibody molecules derived from the sera of the immunized animals.

Monoclonal antibodies, which are homogeneous populations of antibodies to a particular antigen, can be obtained by any

technique which provides for the production of antibody molecules by continuous cell lines in culture. These include, but are not limited to, the hybridoma technique of Kohler and Milstein, (1975, Nature 256:495-497; and U.S. Patent No.

5 4,376,110), the human B-cell hybridoma technique (Kosbor et al., 1983, Immunology Today 4:72; Cole et al., 1983, Proc. Natl. Acad. Sci. USA 80:2026-2030), and the EBV-hybridoma technique (Cole et al., 1985, Monoclonal Antibodies And Cancer Therapy, Alan R. Liss, Inc., pp. 77-96). Such antibodies may
10 be of any immunoglobulin class including IgG, IgM, IgE, IgA, IgD and any subclass thereof. The hybridoma producing the mAb of this invention may be cultivated *in vitro* or *in vivo*. Production of high titers of mAbs *in vivo* makes this the presently preferred method of production.

15 In addition, techniques developed for the production of "chimeric antibodies" (Morrison et al., 1984, Proc. Natl. Acad. Sci. USA 81:6851-6855; Neuberger et al., 1984, Nature, 312:604-608; Takeda et al., 1985, Nature, 314:452-454) by splicing the genes from a mouse antibody molecule of appropriate antigen
20 specificity together with genes from a human antibody molecule of appropriate biological activity can be used (see U.S. Patent Nos. 6,075,181 and 5,877,397 both of which are herein incorporated by reference in their entirety). A chimeric antibody is a molecule in which different portions are derived
25 from different animal species, such as those having a variable region derived from a murine mAb and a human immunoglobulin constant region. Such technologies are described in U.S. Patent Nos. 6,075,181 and 5,877,397 and their respective disclosures which are herein incorporated by reference in their
30 entirety. Also encompassed by the present invention is the use of fully humanized monoclonal antibodies as described in US Patent No. 6,150,584 and respective disclosures which are herein incorporated by reference in their entirety.

Alternatively, techniques described for the production of single chain antibodies (U.S. Patent 4,946,778; Bird, 1988, Science 242:423-426; Huston et al., 1988, Proc. Natl. Acad. Sci. USA 85:5879-5883; and Ward et al., 1989, Nature 341:544-546) can be adapted to produce single chain antibodies against NHS expression products.

Alternatively, techniques described for the production of single chain antibodies (U.S. Patent 4,946,778; Bird, 1988, Science 242:423-426; Huston et al., 1988, Proc. Natl. Acad. Sci. USA 85:5879-5883; and Ward et al., 1989, Nature 341:544-546) can be adapted to produce single chain antibodies against NHS gene products. Single chain antibodies are formed by linking the heavy and light chain fragments of the Fv region via an amino acid bridge, resulting in a single chain polypeptide.

Antibody fragments which recognize specific epitopes may be generated by known techniques. For example, such fragments include, but are not limited to: the F(ab')₂ fragments which can be produced by pepsin digestion of the antibody molecule and the Fab fragments which can be generated by reducing the disulfide bridges of the F(ab')₂ fragments. Alternatively, Fab expression libraries may be constructed (Huse et al., 1989, Science, 246:1275-1281) to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity.

Antibodies to a NHS can, in turn, be utilized to generate anti-idiotypic antibodies that "mimic" a given NHS, using techniques well-known to those skilled in the art. (See, e.g., Greenspan & Bona, 1993, FASEB J 7(5):437-444; and Nissinoff, 1991, J. Immunol. 147(8):2429-2438). For example antibodies which bind to a NHS domain and competitively inhibit the binding of NHS to its cognate receptor can be used to generate anti-idiotypes that "mimic" the NHS and, therefore, bind and activate or neutralize a receptor. Such anti-idiotypic

antibodies or Fab fragments of such anti-idiotypes can be used in therapeutic regimens involving a NHS- mediated pathway.

Additionally given the high degree of relatedness of mammalian NHSs, the presently described knock-out mice (having
5 never seen NHS, and thus never been tolerized to NHS) have a unique utility, as they can be advantageously applied to the generation of antibodies against the disclosed mammalian NHS (i.e., NHS will be immunogenic in NHS knock-out animals).

The present invention is not to be limited in scope by the
10 specific embodiments described herein, which are intended as single illustrations of individual aspects of the invention, and functionally equivalent methods and components are within the scope of the invention. Indeed, various modifications of the invention, in addition to those shown and described herein
15 will become apparent to those skilled in the art from the foregoing description. Such modifications are intended to fall within the scope of the appended claims. All cited publications, patents, and patent applications are herein incorporated by reference in their entirety.